

1           **Low-pressure gas chromatography - ion trap mass spectrometry for the fast**  
2           **determination of polycyclic aromatic hydrocarbons in air samples**

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8  
9           **Abstract**

10           The low-pressure gas chromatography - ion trap mass spectrometry (LPGC-ITMS)  
11           method was investigated to shorten the analysis time for 18 US Environmental Protection  
12           Agency priority listed polycyclic aromatic hydrocarbons (PAHs). Their elution was  
13           optimised with a short, wide-bore column coupled to a deactivated capillary at the inlet end  
14           and with a long, conventional column to compare their analytical performance. The  
15           analytical figures of merit under optimal LPGC-ITMS conditions were determined with  
16           respect to chromatographic separation, S/N ratio, limit of detection and precision. The peak  
17           width at half height of 1.5 s matched the duty cycle of the ITMS. Up to 16 PAHs in the  
18           molecular weight (MW) range of 128-278 Da could be separated in a very short time, i.e.  
19           less than 13 min using LPGC-ITMS, whereas with conventional GC-MS, it took  
20           approximately 40 min. However, LPGC-ITMS has a limited loss of separation power  
21           compared to that of conventional GC-MS due to occurrence of 3 critical pairs for high MW  
22           PAHs. For a practical evaluation, the LPGC-ITMS approach was applied to the  
23           determination of PAHs in gas and aerosol phase samples collected in the ambient air of  
24           Hasselt, Belgium.

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26           **Keywords:** PAHs analysis, fast GC method, LPGC-ITMS, environmental monitoring.

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34 **Introduction**

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36 The application of vacuum column-outlet conditions in a short, wide-bore column is  
37 an attractive way to increase the speed of gas chromatography (GC) analysis, whereas its  
38 compatibility with an ion trap (IT) or a quadrupole mass spectrometer (MS) remains  
39 retained [1-4]. Despite the attractive speed and larger loadability offered by a wide-bore  
40 column operated under vacuum outlet conditions or by low-pressure gas chromatography -  
41 ion trap mass spectrometry (LPGC-ITMS), i.e., when a GC is used with IT detector; there  
42 are only few applications of this technique available [5-8]. Polycyclic aromatic  
43 hydrocarbons (PAHs) are ubiquitous and of major health concern, mainly due to their well-  
44 know carcinogenic and mutagenic properties [9]. Therefore, in the present study, an  
45 application of LPGC-ITMS was elaborated and applied to a very fast determination of  
46 United States Environmental Protection Agency (US EPA) priority listed 18 PAHs [10] in  
47 air samples. The analytical performance of the LPGC-ITMS method was compared to that  
48 of a common GC-MS method with a conventional column. Further, LPGC-ITMS was  
49 applied to the determination of PAHs in ambient aerosol and gas phase samples.

50 **Experimental**

51 ***Instrumentation***

52 A Varian Saturn 2000 IT-MS system was used in combination with a Varian 3800  
53 gas chromatograph (Walnut Creek, CA, USA), equipped with a Varian 1079 universal  
54 injector, being used in splitless mode. Samples were injected with a Varian 8200  
55 autosampler. For the conventional method, a non-polar CP-Sil 8 column (30 m x 0.32 mm  
56 internal diameter (I.D.); film thickness ( $d_f$ ) = 1  $\mu\text{m}$ , Varian Chrompack, Middelburg, The  
57 Netherlands) was applied. For fast GC analysis, a shorter, but wider CP-Sil 8 column (10 m  
58 x 0.53 mm I.D.;  $d_f$  = 1  $\mu\text{m}$ , Varian Chrompack, Middelburg, The Netherlands) was used.  
59 The column was coupled to an uncoated restriction column of 60 cm x 0.1 mm I.D. (Varian  
60 Chrompack) by a single ferrule column connector. Helium (Air Liquide, Liege, Belgium)  
61 was used as a carrier gas for both methods.

62 ***Reagents and standards***

63 The 18 US EPA priority listed PAHs were used either separately or in a mixture of  
64 standard solutions for calibration. These PAHs include naphthalene, acenaphthene,

65 acenaphthylene, fluorene, phenanthrene, anthracene, fluoranthene, pyrene,  
66 benzo[a]anthracene, chrysene, benzo[b]fluoranthene, benzo[k]fluoranthene,  
67 benzo[e]pyrene, benzo[a]pyrene, perylene, indeno[1,2,3-cd]pyrene, dibenz[a,h]anthracene,  
68 benzo[ghi]perylene. Further, a mixture of five perdeuterated PAHs ( $[^2\text{H}_{12}]$ perylene,  
69  $[^2\text{H}_{12}]$ chrysene,  $[^2\text{H}_8]$ naphthalene,  $[^2\text{H}_{10}]$ phenanthrene, and  $[^2\text{H}_8]$ acenaphthene) was used as  
70 internal standards. All chemicals were of analytical reagent grade.

#### 71 ***Samples preparation***

72 Air samples were collected with a high volume sampler (Anderson, OH, USA) from  
73 the ambient air in Hasselt, Belgium, during October-November 2002, as a part of an  
74 environmental monitoring programme of the Flemish Environment Agency (VMM). The  
75 details about the sampling [11] and extraction procedures are described in detail elsewhere  
76 [11, 12]. The extracts were concentrated under a gentle flow of  $\text{N}_2$  up to dryness and were  
77 re-dissolved in 50  $\mu\text{L}$  isooctane. Recovery efficiencies for 18 PAHs were found to be  
78 between 80 and 120 % with the certified reference material of the National Institute of  
79 Standards and Technology (NIST): SRM1650a (Diesel Particulate Matter). The NIST  
80 standard PAHs mixture (SRM 1647d, Schmidt, Amsterdam, The Netherlands) is used for  
81 the calibration of analytical methods and for spiking the samples.

#### 82 **Results and discussion**

##### 83 ***Optimization of LPGC-ITMS conditions***

84 Table 1 lists various GC parameters such as temperature programming rate,  
85 injection temperature, flow rate, and injected volume that were studied to achieve optimal  
86 separation of PAHs with low and high molecular weight (MW). The analyses were  
87 performed by selected ion monitoring (SIM), i.e., measuring the molecular ion of each  
88 compound. Standard mixtures were injected in split and splitless mode. The latter operation  
89 provided increased sensitivity. The injected amounts of individual PAHs between 2.5 and  
90 15 ng by the use of 0.5-3  $\mu\text{L}$  volume from a standard solution with a concentration of 5  
91 ng/ $\mu\text{L}$  had no significant effect on the separation efficiency. The fast temperature  
92 programming (40  $^\circ\text{C}/\text{min}$ ) starting from an initial temperature of 40  $^\circ\text{C}$  onwards gave the  
93 best combination of adequate separation of PAHs and reduction of the analysis time.  
94 Specifically, baseline separation was obtained for the PAHs with MW of 128, 152, 154,  
95 166, 178 and 202 Da.

96 In comparison with GC-MS using a conventional column, most of the mass spectral  
97 parameters such as scan rate, emission current, maximum ionisation time and multiplier  
98 offset did not require major changes for LPGC-ITMS (Table 1). The scanrate was  
99 increased to 3 scan/s to improve the resolution of the chromatographic peak profiles.

#### 100 ***Comparison of conventional GC-MS and LPGC-ITMS for the determination of PAHs***

101 The analysis of a standard PAH mixture was carried out using conventional GC-MS  
102 and LPGC-ITMS methods under optimized conditions. Fig. 1 shows the corresponding  
103 total ion current chromatograms. The elution on the conventional column was optimised  
104 following the US EPA Method TO-13A [10], resulting in a total analysis time of 40 min. In  
105 contrast, the use of LPGC-ITMS allowed the elution time to be reduced to less than 13 min.  
106 A further advantage of the method was the low elution temperature that in turn,  
107 significantly reduced the background, due to column bleeding. For a detailed evaluation of  
108 the chromatographic separation obtained on conventional and LPGC-ITMS columns, the  
109 mass chromatograms of individual PAHs were compared. In general, the peak width at half  
110 height ( $W_h$ ) was found to be narrower with the LPGC-ITMS than with the conventional  
111 method, e.g., yielding for fluoranthene the values of 1.2 and 2.1 s, respectively.  
112 Furthermore, the LPGC-ITMS method fully retained the chromatographic separation  
113 efficiency of GC-MS with a conventional column for PAHs in the MW range of 128-202  
114 Da. However, the reduction in the analysis time sacrificed some resolution for PAH analogs  
115 with MW between 228 and 252 Da, i.e. occurrence of 3 critical pairs of PAHs, two of them  
116 separated below the half height. Since the quantitation was performed in SIM with other  
117 target ions, it was possible to identify and quantify them based on their separation near half  
118 height and the slight difference in their retention times. Therefore, we were able to  
119 determine 16 PAHs except for benzo[b]fluoranthene and benzo[k]fluoranthene, which were  
120 quantified in total. Interestingly, LPGC-ITMS allows high MW PAHs (276, 278 Da) to be  
121 eluted with adequate separation, but much faster than in the case of a conventional column  
122 (peaks 16, 17 on Fig. 1). The calibration curves based on diluted standards showed  
123 correlation coefficients better than 0.999 for each PAH. Table 2 lists the separation power  
124 of the two methods expressed as the numbers of theoretical plates ( $n$ ) by the formula of  $n =$   
125  $5.545 (t_r/W_h)^2$ , where  $t_r$  is the retention time of the peak.

126 Analysis of serial dilutions of PAH standards shows that the absolute limits of  
127 detection (LOD) for the MW range from 128 to 202 Da varied from 50 pg to 140 pg, and

128 are comparable to those of the conventional method (65 to 120 pg). Unfortunately, this  
129 observation does not extend to the PAH analogues between 228-252 Da, because the loss in  
130 chromatographic separation comes together with a significant reduction in the detection  
131 capabilities. The LOD in this range varied from 70 to 150 pg, except for  
132 benzo[a]anthracene (650 pg) and perylene (1000 pg). In contrast, the high MW PAHs (276  
133 and 278 Da) were detected with proper resolution and with a similar sensitivity to that of  
134 the conventional method. A LOD of 320 pg was reported for indeno[1,2,3-cd]pyrene and  
135 benzo[ghi]perylene, while it was 1650 pg for dibenz[a,h]anthracene. The comparative LOD  
136 values in conventional columns lies in the range of 750 pg on column. The above LOD data  
137 are also comparable with those reported by Sheu et al. [13] and Sofuoglu et al. [14].  
138 However, it has to be noted that the larger loadability of a wide-bore capillary column may  
139 further improve the LOD.

140 Additional parameters for a trace analysis method, the precision and linear range of  
141 quantitation were observed to be adequate for quantitation of PAHs. Repeated (18)  
142 injections of a PAHs mixture with a concentration of  $2.5 \text{ ng } \mu\text{L}^{-1}$  shows relative standard  
143 deviations of 5-15 %. This analytical performance of LPGC-ITMS is considered to be  
144 adequate for a fast routine monitoring of PAH levels in environmental samples.

#### 145 *Practical evaluation of the LPGC-ITMS*

146 The performance of the LPGC-ITMS method was evaluated by the analysis of air samples  
147 collected from Hasselt (Belgium) near a highway. During the monitoring period the daily  
148 average concentrations of total PAHs ( $\Sigma 18$ ) varied from  $24 \pm 4 \text{ ng m}^{-3}$  to  $33 \pm 4 \text{ ng m}^{-3}$  in gas  
149 phase samples, whereas it ranged from  $6.6 \pm 1.3 \text{ ng m}^{-3}$  to  $8.5 \pm 0.6 \text{ ng m}^{-3}$  for the aerosol.  
150 The daily levels of individual PAHs in the gas phase samples varied from below the LOD  
151 to  $19 \text{ ng m}^{-3}$ , and showed the prevalence of low and medium MW PAHs, such as  
152 phenanthrene,  $16 \pm 3.1 \text{ ng m}^{-3}$ ; fluorene,  $3.3 \pm 0.7 \text{ ng m}^{-3}$ ; fluoranthene,  $2.4 \pm 1.0 \text{ ng m}^{-3}$ ; and  
153 pyrene,  $2.0 \pm 1.0 \text{ ng m}^{-3}$ . The particulate phase reflected the occurrence of predominantly  
154 high MW PAHs (dibenz[a,h]anthracene,  $2.5 \pm 1.9 \text{ ng m}^{-3}$ ; benzo[ghi]perylene,  $2.2 \pm 1.5 \text{ ng m}^{-3}$ ;  
155 and indeno[1,2,3-cd]pyrene,  $1.9 \pm 1.0 \text{ ng m}^{-3}$ ). The daily individual PAH concentrations  
156 ranged up to  $3.9 \text{ ng m}^{-3}$ , whereas the more volatile PAHs were found to be at lower levels  
157 in the particulate matter.

#### 158 **Conclusion**

159           Compared to the conventional GC-MS methodology, LPGC-ITMS provides a  
160 valuable alternative for a fast analysis of PAHs in air samples. This new method allows the  
161 analysis time to be reduced by a factor of three with the preservation of the  
162 chromatographic resolution for the low MW PAHs that are prevalent in the gas phase  
163 samples of the ambient air. The loss of separation power for high MW PAHs is an  
164 acceptable shortcoming, when the increased sample throughput is taken into account and  
165 when information on concentration of all individual (e.g. 18 US EPA) PAHs is not  
166 required. Furthermore, LPGC-ITMS is an affordable method that can be readily  
167 implemented on current GC-MS systems without any major change in the instrumental  
168 configuration.

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**Table 1. Chromatographic and mass spectrometric parameters studied for the optimization of PAH analysis using the LPGC-ITMS and conventional GC-MS**

Parameter	Studied range	Optimised values	
		LP-GC	Conventional
Temperature programming rate ( $^{\circ}\text{C min}^{-1}$ )	0-60	<i>a</i>	<i>b</i>
Injector temperature ( $^{\circ}\text{C}$ )	200-300	290	290
Gas flow rate ( $\text{ml min}^{-1}$ )	1-5	1.2	2.0
Injection volume ( $\mu\text{l}$ )	0.5-10	1	1
Transfer line temperature ( $^{\circ}\text{C}$ )	200-370	230	240
Ion trap temperature ( $^{\circ}\text{C}$ )	200-260	220	230
Scan time ( $\text{s scan}^{-1}$ )	0.22-0.8	0.35	0.50
Multiplier offset (V)	-20 to +50	0	0
Emission current ( $\mu\text{A}$ )	20-60	50	50
Max. ionization time (ms)	10 -50	30	35

205 <sup>a</sup> [  $40^{\circ}\text{C}$  (1 min)  $\rightarrow 120^{\circ}\text{C}$ ( $40^{\circ}\text{C min}^{-1}$ )  $\rightarrow 260^{\circ}\text{C}$ ( $15^{\circ}\text{C min}^{-1}$ ) ]

206 <sup>b</sup> [  $70^{\circ}\text{C}$  (5 min)  $\rightarrow 120^{\circ}\text{C}$ ( $15^{\circ}\text{C min}^{-1}$ )  $\rightarrow 300^{\circ}\text{C}$ ( $5^{\circ}\text{C min}^{-1}$ ) ]

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**Table 2. Comparison of theoretical plate numbers calculated for PAH analysis with conventional GC-MS and LPGC-ITMS methods**

Compounds	Theoretical Plate Numbers*	
	Conventional GC-MS	LPGC-ITMS
Naphthalene	1711	40
Acenaphthylene	1853	143
Phenanthrene	1277	204
Anthracene	2461	456
Fluoranthene	1790	486
Pyrene	1800	565
Benzo[e]pyrene	930	646
Benzo[a]pyrene	884	563
Dibenz[a,h]anthracene	360	466

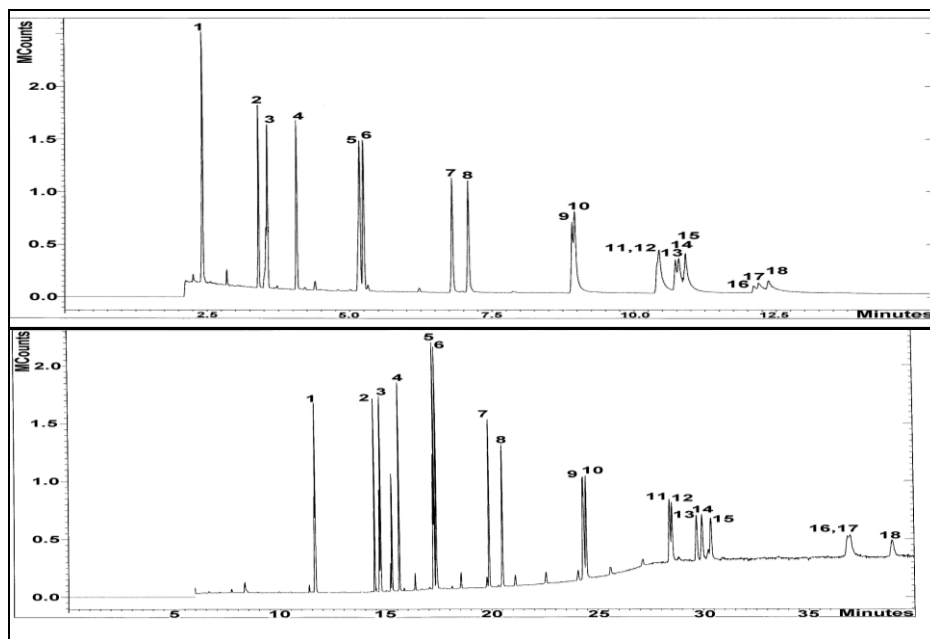
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Figure 1.

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**Figure caption**

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Figure 1. Comparison of the analysis of a PAH mixture using LPGC-ITMS (top) and CP-Sil 8 conventional column (bottom). Naphthalene (1), acenaphthene (2), acenaphthylene (3), fluorene (4), phenanthrene (5), anthracene (6), fluoranthene (7), pyrene (8), benzo[a]anthracene (9), chrysene (10), benzo[b]fluoranthene (11), benzo[k]fluoranthene (12), benzo[e]pyrene (13), benzo[a]pyrene (14), perylene (15), indeno[1,2,3-cd]pyrene (16), dibenz[a,h]anthracene (17), benzo[ghi]perylene (18).